

Research Article

Determination of three active compounds of methanolic extract of *Schisandra chinensis* by RP-HPLC

BAI Jing^{1,2}¹Research Center of Life Sciences and Environmental Sciences, Harbin University of Commerce, China²Research Center of Nature Drug Engineering for Anti-tumor, Ministry of Education, Harbin 150076, China

*Corresponding Author:

BAI Jing

Email: baijing0308@163.com

Abstract: To establish a method for determination of contents of schizandrol A, deoxyschisandrin and schisandrin B in *Schisandra chinensis* (Turcz.) Baill. An RP-HPLC method was developed. A Kromasil C₁₈ column (250 mm×4.6 mm, 5 μm) was used, with the mobile phase of acetonitrile-water by gradient elution, the flow rate was 1.0 mL·min⁻¹ and the UV absorbance was monitored at 218 nm. The linear range of schizandrol A, deoxyschisandrin, schisandrin B were 19.92~398.4 μg·mL⁻¹, 6.24~124.8 μg·mL⁻¹, 12.5~250 μg·mL⁻¹, the mean recovery were 99.5%, 98.6%, 98.5%. The method is suitable for the quality control of *Schisandra chinensis* (Turcz.) Baill.

Keywords: schizandrol A; deoxyschisandrin; schisandrin B; RP-HPLC

INTRODUCTION

Fructus Schisandrae chinensis, originated from the dried ripe fruits of *Schisandra chinensis* (Turcz.) Baill. was ranked as high-grade herbal drug in ancient medical book 'Shen Nong Bencao Classics'. Lignans were the most abundant and active components derivative isolated from the fruit of *Schisandra chinensis*, which has demonstrated wide range of pharmacological activities including; hepatoprotective and neuroprotective effects, antioxidant, anti-inflammatory, immunomodulating effects and so on [1-10]. In the past few years, chromatographic techniques has been successfully applied to determine the contents of bioactive components in herbal medicines [11, 12].

In our study, *Schisandra chinensis* were extracted by water, 75% ethanol and methanol respectively in order to screening of active fraction. The result of the sedative and hypnotic experiments indicated that the methanol extract showed the highest pharmacological activities than that of the other extracts. The results showed that the methanol extract of *Schisandra chinensis* can shorten sleep latency significantly, increase sleeping time. Herein, we developed methods for a systematic identification of active fractions of *Schisandra chinensis*. In this study, a simple, reliable and sensitive analysis method by high-performance liquid chromatography coupled with diode array detection was developed for quantitative determination of the three active compounds in Fructus Schisandrae chinensis.

MATERIALS AND METHODS

Instrumentation

HPLC analyses were performed using a Shimadzu RP-HPLC system (Shimadzu Corporation, Japan) consisting of an SCL-10AVP system controller, LC-10ATvp infusion pumps, SPD-10Avp UV detector, ANASTAR chromatography workstation. Mettler Analytical Balance AE240 (Mettler-Toledo Instruments Co., Ltd.)

Chemical reagents

Schizandrol A (lot number: 0857-200304), deoxyschisandrin (lot number: 0764-200107), schisandrin B (lot number: 0765-200205). Standard reference were all obtained from National Institute for the Control of Pharmaceutical and Biological Products, China. HPLC-grade acetonitrile were obtained from Fisher Scientific (Pittsburgh, PA), ultra pure water, *Schisandra chinensis* was collected from Heilongjiang, China.

Preparation of Sample solution

About 1 g of the sample was accurately weighed and ultrasonic extracted by 50 ml menthol for 30 min, make up the weight loss, shake, and the supernatant filtered (0.45 μm), made for the test solution.

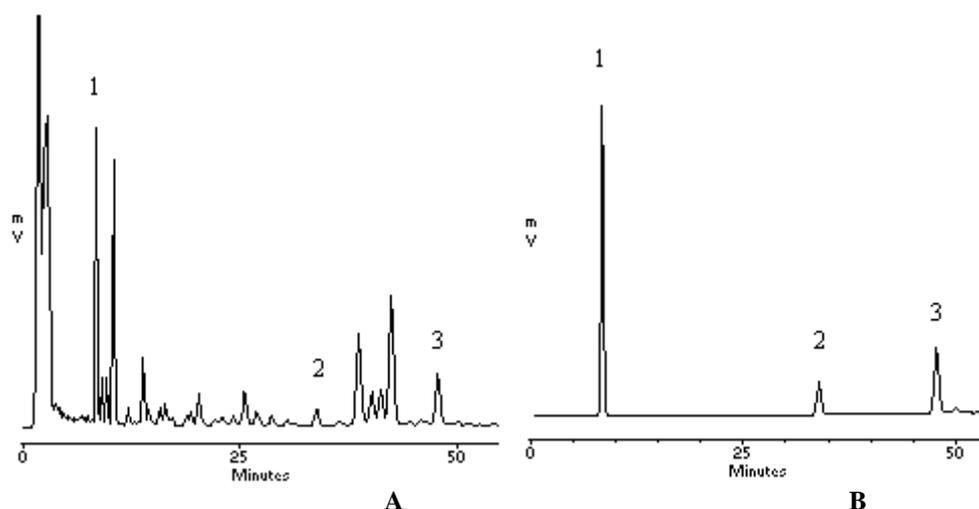
Preparation of mixed standard solution

Reference substance schizandrol A, deoxyschisandrin and schisandrin B were accurately weighed and dissolved in 50 ml of methanol as a stock solution. The final concentration of the standard solution were 199.2, 62.4 and 125 μg/ml, respectively

Chromatographic Conditions

Column was Kromasil C18 (5 μm , 4.6 mm \times 250 mm); A binary gradient elution system consisting of acetonitrile (A) and water (B) was used with the

following gradient program: 0-55 min, 70% B-55%B; detection wavelength was set at 218 nm and the flow rate was 1.0 mL/min. The injection volume was 20 μl and the column temperature was maintained at 30°C.



**Fig-1:HPLC chromatograms of *Schisandra chinensis* fruits (A), reference substances (B)
1- schizandrol A 2- deoxyschisandrin 3- schisandrin B**

RESULTS AND DISCUSSION

Linearity

The calibration curve were obtained by plotting the concentration($\mu\text{g/ml}$) as the x-axis versus peak areas on Y-axis and regression equations were computed. Six

concentrations of the four Standards were analyzed. Calibration curves of all analytes showed good linearity over a wide concentration range. The test results were given in Table-1.

Table-1: Result of regression analysis on calibration curves

Compd	Regression equation	r^2	Linear range ($\mu\text{g/mL}$)
schizandrol A	$Y_1=5.587 \times 10^6 X+1.212 \times 10^6$	0.9991	19.92~398.4
deoxyschisandrin	$Y_2=3.756 \times 10^7 X+2.693 \times 10^6$	0.9996	6.24~124.8
schisandrin B	$Y_3=1.777 \times 10^7 X-5.550 \times 10^5$	0.9994	12.5~ 250

Precision

The stock solutions of reference substances were measured for five times under the same analytical conditions in a day. the relative standard deviation (RSD) values of retention time were all lower than 0.3%, while the RSD values of peak area were 0.80%, 0.47% and 0.62%, respectively. The results showed that the assay had a good intra-day precision.

Stability

For the stability testing, sample solution was stored at ambient temperature (25°C) for 24h. The same real sample was re-analyzed after 12 and 24h time intervals and compared against fresh sample. The RSD values of the stability was less than 2.0%. It is illustrated that the solution was found to be stable within 24h during assay determination.

Accuracy

The accuracy of the method was determined by means of recovery experiments. Particular amount of

the authentic standards were added into the pre-analyzed sample solution at three levels (80%, 100% and 120%, three replicates of each concentration). The mixed samples were analyzed using the developed HPLC method mentioned above. The average recoveries of schizandrol A, deoxyschisandrin and schisandrin B were 99.5%, 98.6% and 98.5% respectively, and RSDs of the above three compounds were 1.7% (n=9), 1.9% (n=9) and 1.6% (n=9) respectively. It was confirmed from recovery results that this developed method is highly accurate for the determination of these components.

Sample analysis

The validated HPLC method was applied to the simultaneous determination of three active components in herbs. The contents of schizandrol A, deoxyschisandrin and schisandrin B were calculated by external standard method (Table-2).

Table-2 Determination results of samples (w%, n=3)

No	schizandrol A	deoxyschizandrin	schisandrin B
1	0.41	0.09	0.18
2	0.48	0.11	0.20
3	0.52	0.13	0.26
4	0.61	0.15	0.29
5	0.49	0.12	0.22

CONCLUSION

Under above optimized chromatographic conditions, 3 chromatographic peak were well separated. The convenient and high specific method could be used to identify and evaluate the quality of *Schisandra chinensis*. The developed method also lays a solid foundation for further comprehensive study on the pharmacodynamic-pharmacokinetic effects of *Schisandra chinensis*.

PREFERENCES

- Xu, X., Zhou, X., Zhou, X. W., Zhang Z., Liao, M. G., Gao, Q., & Luo, H. M. (2012). Schizandrin prevents dexamethasone-induced cognitive deficits. *Neuroscience Bulletin*, 28 (5), 532-540.
- Lee, T.H., Jung, C.H., & Lee, D.H. (2012). Neuroprotective effects of Schisandrin B against transient focal cerebral ischemia in Sprague-Dawley rats. *Food and Chemical Toxicology*, 50 (12), 4239-4245.
- Pan, S.Y., Dong, H., Zhao, X.Y., Xiang, C.J., Fang, H.Y., Fong, W.F., Yu, Z.L., & Ko, K.M. (2008). Schisandrin B from *Schisandra chinensis* reduces hepatic lipid contents in hypercholesterolaemic mice. *Journal of Pharmacy and Pharmacology*, 60 (3), 399-403.
- Li, L., Zhang, T., Zhou, L., Zhou, L., Xing, G., Chen, Y., & Xin, Y. (2013). Schisandrin B attenuates acetaminophen-induced hepatic injury through overexpression of heat shock protein 27 and 70 in mice. *Journal of Gastroenterology and Hepatology*, 29 (3), 640-647.
- Checker, R., Patwardhan, R.S., Sharma, D., Menon, J., Thoh, M., Bhilwade, H.N., Konishi, T., & Sandur, S.K. (2012). Schisandrin B exhibits anti-inflammatory activity through modulation of the redox-sensitive transcription factors Nrf2 and NF- κ B. *Free Radical Biology and Medicine*, 53 (7), 1421-1430.
- Chang, R., Li, Y., Yang, X., Yue, Y., Dou, L., Wang, Y., Zhang, W., & Li, X. (2013). Protective role of deoxyschizandrin and schisantherin A against myocardial ischemia-reperfusion injury in rats. *PLoS One*, 8 (4), e61590.
- Guo, C.X., Deng, S., Yin, J.Y., Liu, Z.Q., Zhang, W., & Zhou, H.H. (2015). Schisandrin A and B induce organic anion transporting polypeptide 1B1 transporter activity. *Pharmazie*, 70(1), 29-32.
- Wu, J., Cao, Y., Zhang, Y., Liu, Y., Hong, J.Y., Zhu, L., Ge, G., & Yang, L. (2014). Deoxyschizandrin, a naturally occurring lignan, is a specific probe substrate of human cytochrome P450 3A. *Drug Metab Dispos*, 42(1), 94-104.
- Gu, B.H., Minh, N.V., Lee, S.H., Lim, S.W., Lee, Y.M., Lee, K.S., & Kim, D.K. (2010). Deoxyschizandrin inhibits H₂O₂-induced apoptotic cell death in intestinal epithelial cells through nuclear factor-kappaB. *International Journal of Molecular Medicine*, 26(3), 401-406.
- Ba, Q., Cui, C., Wen, L., Feng, S., Zhou, J., & Yang, K. (2015). Schisandrin B shows neuroprotective effect in 6-OHDA-induced Parkinson's disease via inhibiting the negative modulation of miR-34a on Nrf2 pathway. *Biomed Pharmacother*, 75, 165-172.
- Wu, X.D., Chen, H.G., Zhou, X., Huang, Y., Hu, E.M., Jiang, Z.M., Zhao, C., Gong, X.J., & Deng, Q.F. (2015). Studies on Chromatographic Fingerprint and Fingerprinting Profile-Efficacy Relationship of *Saxifraga stolonifera* Meerb. *Molecules*, 20 (12), 22781-22798.
- Yi, J., Wu, J.G., & Wu, J.Y. (2016). Quality evaluation of the leaves of *Magnolia officinalis* var. *biloba* using high-performance liquid chromatography fingerprint analysis of phenolic compounds. *Journal of Separation Science*, 39 (4), 784-792.