

## Antibacterial Activity of Methanolic Fruit Extract of *Randia dumetorum* Lamk against Ocular Pathogens

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### Original Research Article

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#### Article History

Received: 08.06.2018

Accepted: 19.06.2018

Published: 30.06.2018

#### DOI:

10.21276/sjmeps.2018.4.6.9



**Abstract:** Ocular diseases have been documented as the most common health problems universally. Most of the chemicals and synthetic drugs currently in use have marked side effects. Hence, there has been an ideal shift from the use of modern drugs to the age-old herbs. *Randia dumetorum* Lamk is one such important plant with various established medicinal properties. The aim of the present study was to evaluate the preliminary antibacterial activity of methanolic extract of *Randia dumetorum* Lamk. (*Xeromphis spinosa* Thumb) Against common ocular pathogens such as *Serratia marcescens*, *S. agalactiae*, *Corynebacterium macbinleys* and *Propionibacterium acnes*. Methanolic extract of the dried fruits of the plant was prepared. Different concentrations of the dried fruit extracts (*R. dumetorum*) were transferred to the nutrient agar plates, which had been previously inoculated with the test microorganisms. The plates were incubated at 37°C for 24 h in an incubator and the zones of inhibition were measured using well diffusion method. The extract showed potential antibacterial properties comparable with that of the standard streptomycin against the organisms tested. The methanol extract of *R. dumetorum* displayed a concentration related antibacterial activity. The results showed that the inhibition of the bacterial growth was more pronounced on *Corynebacterium macbinleys* as compared to the other tested organisms.

**Keywords:** *Randia dumetorum* Lamk, antibacterial activity, *Serratia marcescens*, *S. agalactiae*, *Corynebacterium macbinleys* and *Propionibacterium acnes*.

## INTRODUCTION

Ocular infections can cause damage to the structures of eye, which can lead to reduced vision and even blindness if left untreated [1]. The cause of ocular infections can be bacteria, fungi, parasites and viruses. Gram positive bacteria are most common pathogens infecting all the tissues of the eye.

The most frequently effected part of the eye is conjunctiva, lid and cornea which are the external parts of the eye [2]. External bacterial infections of eye are usually localized but may frequently spread to the adjacent tissue due to some predisposing factors such as during trauma, previous surgery, ocular surface disease; contact lens wear etc., systemic diseases and immunosuppression may alter the defense mechanisms of the outer eye and permit bacteria to spread [3]. Infections of the external eye account for a significant percentage of ocular inflammations, some of which lead to visual losses as a result of corneal involvement [4].

Many chemicals and synthetic drugs have proven to be effective in the prevention of these

diseases, but they also have marked side effects. In recent times, there has been a marked shift toward herbal cures because of the pronounced cumulative and irreversible reactions of modern drugs [3,4].

*Randia dumetorum lamk* is a species of plant in the Rubiaceae family, commonly known as Mainphal, Mindhal. It is found in waste places & jungles all over India, extending northwest to the Bias River & ascending to outer Himalaya to 4000 ft [5].

Ripe fruits of the plant contain glycosides, randioside A, mollisidial triterpenoid glycosides and randianin, six saponins dumetoronins A to F [6, 7]. The fruit is reported to have various pharmacological actions such as aphrodisiac, emetic, purgative, carminative, antipyretic, cures abscess, ulcers, inflammations, wounds, tumors, skin diseases and have antibacterial activity. The pulp of fruit is believed to have anthelmintic properties, and also used as an abortifacient in folklore remedy. The bark is astringent and is given in cases of diarrhoea and dysentery [8].

Various studies report that the Methanolic extract of *Randia dumetorum Lamk* has shown antibacterial activity against *Staphylococcus aureus*, *Staphylococcus epidermidis*, *Bacillus cereus*, *Bacillus subtilis*, *Escherichia coli* and *Salmonella typhi* [9]. Very few studies have been conducted on the antibacterial effect of *Randia dumetorum Lamk* extracts against ocular pathogens. Therefore, the aim of this study is to evaluate the *in vitro* antibacterial activity of *R.dumetorum* against some selected ocular pathogens.

## MATERIALS AND METHODS

### Plant collection and authentication

*Randia dumetorum* seeds were obtained from local market and authenticated by Head of Botany department, Malla Reddy college of Pharmacy, Maisammaguda, Telangana. A voucher specimen has been deposited in the museum of department of Pharmacognosy, M.R.C.P Telangana. Voucher specimen (PH-709) was also deposited in the herbarium of Pharmacy Department of M.R.C.P Telangana.

### Plant preparation and extraction

The fruits were dried in sunlight and reduced to a coarse powder. Then the powder was subjected to Soxhlet extraction with methanol for 72 hours at a temperature of 50-60°C. The extract was concentrated and the solvent was completely removed. They were freeze dried and stored in the vacuum desiccator until further use. Preliminary phytochemical screening was carried out to identify the chemical constituents [10].

### Microorganism

Pure cultures of *Serratia marcescens*, *S. agalactiae*, *Corynebacterium macbinleys* and *Propionibacterium acnes* were obtained from MTCC, Institute of Microbial Technology, and Chandigarh.

### Preparation of inoculums

The suspension of all organisms were prepared by inoculating a single colony of the strain in 20 ml of sterile nutrient broth and incubated at 37°C for 24 hours. The suspension is adjusted such that it contained approximately  $1 \times 10^6$  cells/ml. It was obtained by calculating the cells by Neubers chamber. Nutrient agar (HiMedia) was prepared for the study.

### Controls for the test

The controls were needed to confirm all the necessary nutritional conditions were suitable for the growth of microorganism and for absence of inhibitory substance in the medium. The positive controls were observed by streaking the organism on agar plates for observing morphology of colonies. Any contamination during the assay was ruled out by keeping the negative control. This was checked by adding the sterile saline and observing for growth as a contamination. The results indicated that the medium was free from contamination.

### Culture medium

Readymade dehydrated medium supplied by Hi Media was used for testing the antimicrobial activity of plant extracts. The dehydrated medium was prepared by incorporating 13grams of NA in 100 ml of distilled water and heated to boiling to dissolve the medium completely. The medium was distributed into clean glass tubes and plugged with cotton and sterilized by autoclaving at 15 lb/sq. inch pressure at 121°C for 20 min.

### Antimicrobial Agent

Streptomycin 50mg/ml was used as the reference standard was procured as gift sample from Hindustan Antibiotics Ltd., Pune.

### Determination of MIC (minimum inhibitory concentration)

Broth dilution method was followed for the determination of minimum inhibitory concentration of the extract. Fresh amount of the nutrient broth was prepared and sterilized by autoclaving. 20 ml of the sterilized nutrient broth was transferred to the test tubes. Measured amount of the extract was added in to the test tubes containing nutrient broth in such a way that the final concentration per ml was 0 (control), 5,15,25,50 and 100mg. Loopful of test microorganism was incorporated into the test tubes and incubated at 37°C for 24hrs. After completion of the incubation period, the tubes were checked for the growth. Growth in the liquid cultures was seen in the form of turbidity. Tubes showing growth was denoted by '+' and '-' for absence of growth.

### Determination of zone of inhibition by cup plate method [10]

The antimicrobial activity of the methanolic extract was determined by well diffusion method. 20ml of sterile nutrient agar medium was transferred into the sterile petri-dishes and allowed to solidify. The petri dishes were incubated at 37°C for 24 hours to check for the sterility. The medium was seeded with the organisms by pour plate method using sterile top agar layer (4 ml) containing 1 ml of the culture. Wells were made on the medium using sterile borer. Dried methanolic extract of fruits of *Randia dumetorum* was dissolved in Dimethyl sulfoxide (DMSO) to obtain different concentration (50, 100 and 150 mg/ml) and sterilized by filtration through a Whatman filter paper no. 1, and 0.1 ml of the different concentrations of extract were added to the respective bores. 0.1ml of streptomycin was taken as standard reference. The plates were incubated overnight at 37°C with appropriate positive and negative controls. The petri-dishes were kept in refrigerator at 4°C for ½ hour for diffusion. After diffusion the petri-dishes were incubated at 37°C for 24 hours and zone of inhibition were observed and measured. Dimethyl sulfoxide was used as the control.

## RESULTS AND DISCUSSION

**Table-1: Determination of MIC of methanolic fruit extract of *R. dumetorum* against different bacteria**

Name of the bacteria	Growth in nutrient broth containing different concentration of extract in mg/ml					
	0	5	15	25	50	100
<i>Serratia marcescens</i>	+	+	+	-	-	-
<i>S. agalactiae</i>	+	+	+	-	-	-
<i>Corynebacterium macbinleys</i>	+	+	-	-	-	-
<i>Propionibacterium acnes</i>	+	+	+	-	-	-

'0' – Control (without extract); '+' – Growth; '-' – No growth

**Table-2: Antibacterial activity of Streptomycin and methanolic fruit extract**

Micro organism	Zone of inhibition in mm			
	Extract Conc. mg/ml			Conc. Streptomycin mg/ml
	50	100	150	0.5
<i>Serratia marcescens</i>	12 ± 0.42	13 ± 0.53	14 ± 0.17	18 ± 0.31
<i>S. agalactiae</i>	12 ± 0.51	13 ± 0.18	15 ± 0.57	18 ± 0.42
<i>Corynebacterium macbinleys</i>	14 ± 0.73	16 ± 0.41	21 ± 0.61	23 ± 0.15
<i>Propionibacterium acnes</i>	12 ± 0.42	13 ± 0.53	14 ± 0.17	18 ± 0.31

Plant based medications have been used from many ages. Our ancestors had been known to use herbal drugs to treat and alleviate illnesses that affect the human body. Medicinal plants are today being once again preferred because of the various reasons such as their easy availability, negligible side effects, low cost of treatment, and their effectiveness. The present study proved that *R. dumetorum* possesses a significant antibacterial activity against *Serratia marcescens*, *S. agalactiae*, *Corynebacterium macbinleys* and *Propionibacterium acnes*, which are the causative organisms playing a major role in the pathogenesis of eye infections.

The antibacterial activity of the extract was confirmed by observing zone of inhibition around the well containing test solution after specified incubation period. Absence of bacterial growth in the negative control plate confirmed the sterility of the medium. The remaining plates were examined for the presence or absence of growth. The positive control without *R. dumetorum* extract was checked to ensure that each test strain was capable of exhibiting adequate growth in the medium. The negative control was checked for the absence of growth there by indicating the sterility of the medium. In reading the end points, a faint haze of growth of a single colony was evident for antimicrobial activity. A dense film of growth or more than one colony was considered as evidence that the plant extract failed to inhibit the growth.

The results of the MIC as well as well diffusion are summarized in Table. 1 and 2. The largest zone of inhibition of methanol extracts was found to be 21 mm against *Corynebacterium macbinleys* Second largest zone was observed against *S. agalactiae* i.e. 15 mm at 150mg/ml. Streptomycin was used as the standard drug to check the efficacy of the test compound. At 0.5 mg/ml concentration, it showed a

zone of inhibition of 23 ± 0.15 against *Corynebacterium macbinleys* and 18 mm against remaining organisms. Though the plant extract was found to be effective at a higher concentration and volume than streptomycin, it showed a marked antibacterial activity and should be considered to replace the synthetic drugs and chemicals because of their irreversible side effects.

Any antimicrobial agent is considered effective, given the size of inhibition zone produced by it measures 2 mm or more. In the present study, the minimum zone of inhibitions obtained were 12 mm and 14 mm for methanolic extracts of *R. dumetorum*, respectively. It has proved to have potent antibacterial property.

The result showed that the methanolic fruit extract of *R. dumetorum* displayed concentration dependent antibacterial activities. It indicated that *R. dumetorum* exhibited antibacterial activity towards all four ocular pathogens. The highest antibacterial activity was found towards *Corynebacterium macbinleys*, while it was less active against the remaining. The compounds responsible for this antimicrobial property were not investigated. However preliminary phytochemical analysis of the methanolic extract revealed the presence of phytosterol, polyphenol, saponins, flavonoids and carbohydrates [11]. The antimicrobial potency of the plant may be attributed to the single or combined effect of the above mentioned chemical groups. The methanolic fruit extract of *R. dumetorum* had impressive antibacterial activity and could lead to the discovery of new antibacterial agents. This becomes more relevant as the current drugs in use are fast losing effectiveness due to their irreversible side effects.

## CONCLUSION

In the present study methanolic extract of *Randia dumetorum Lamk* showed good activity against the ocular pathogens. Further studies must be conducted for the separation of the active components of the extract and to assess its safety levels.

## Conflict of interest

The authors declare no conflict of interest.

## Authors' Declaration

The authors hereby declare that the work presented in this article is original and that any liability for claims relating to the content of this article will be borne by them.

## ACKNOWLEDGEMENTS

The authors wish to thank and appreciate the management of Malla Reddy College of Pharmacy College for providing necessary facilities to carry out the research work.

## REFERENCES

1. Sherwal, B. L., & Verma, A. K. (2008). Epidemiology of Ocular Infection Due to Bacteria and Fungus—A Prospective Study.
2. Tesfaye, T., Beyene, G., Gelaw, Y., Bekele, S., & Saravanan, M. (2013). Bacterial profile and antimicrobial susceptibility pattern of external ocular infections in Jimma University Specialized Hospital, Southwest Ethiopia. *American Journal of Infectious Diseases and Microbiology*, 1(1), 13-20.
3. Sharma, S. (1988). Ocular Microbiology. 1st ed. Madurai: Arvind Eye Hospital Publication.
4. Sharma, S. H. I. V. A., Devine, W., Anderson, R. H., & Zuberbuhler, J. R. (1988). The determination of atrial arrangement by examination of appendage morphology in 1842 heart specimens. *Heart*, 60(3), 227-231.
5. Prabhat, A., & Navneet, C. A. (2010). Evaluation of antimicrobial activity of six medicinal plants against dental pathogens. *Report opinion*, 2(6), 37-42.
6. Srivastav, S., Singh, P., Mishra, G., Jha, K. K., & Khosa, R. L. (2011). *Achyranthes aspera*-An important medicinal plant: A review. *J Nat Prod Plant Resour*, 1(1), 1-14.
7. Nandkarni, A. K. (1982). *Indian Materia Medica Popular Prakashan Bombay Vol I&II* (reprinted).
8. Agarwal, S. S. (1999). Immunomodulation: A review of studies on Indian medicinal plants and synthetic peptides, Part 1: medicinal plants. In *Proc. Ind. Natl. Sci. Acad.* (Vol. 65, No. 3, pp. 79-204).
9. Kirtikar, K. R., & Basu, B. D. (1991). *Indian Med. Plants*, 3, 2274-2277.
10. Chopra, R. N., & Nayar, S. L. (1956). *Glossary of Indian medicinal plants*. Council of Scientific And Industrial Research; New Delhi.
11. Dharmishtha, M., & Falguni, G. (2009). Antibacterial activity of methanolic fruit extract of *Randia dumetorum lamk*. *International Journal of PharmTech Research*, 1(3), 679-681.
12. Kokate, C. K. (1986). Preliminary phytochemical analysis. *Practical Pharmacognosy. 1st ed. New Delhi: Vallabh Prakashan*, 111.
13. Pelczar M.J. and Reid J.D. (1974). *Microbiology*, Tata Mcgraw Hill, New Delhi. 473.